

# Green Synthesis and Characterization of Zinc Oxide Nanoparticles Using *Cyperus pangorei* Grass Extract and its Antibacterial Activity



Rohan Gavankar\*<sup>1</sup>, Deepa Verma<sup>1</sup>, Ankita S. Tekade<sup>2</sup>, and Virendra Sangode\*<sup>3</sup>

<sup>1</sup>Department of Botany, Vishnu Waman Thakur Charitable Trust's Bhaskar Waman Thakur College of Science. Yashvant Keshav Patil College of Commerce, Vidhya Dayanand Patil College of Arts, VIVA COLLEGE, Virar West, India

<sup>2</sup>Department of Zoology, Shivaji Science College, Pauni, Maharashtra, India

<sup>3</sup>Department of Zoology, Manoharbai Patel College of Arts Commerce and Science Sadak Arjuni District Gondia, India

## ABSTRACT

Green synthesis of nanoparticles using plant-derived biomolecules has emerged as an eco-friendly and sustainable approach in nanobiotechnology. In the present study, zinc oxide nanoparticles (ZnO NPs) were biosynthesized using aqueous leaf extract of *Cyperus pangorei*, a biologically important grass species rich in phytochemicals such as flavonoids, phenolics, alkaloids, and terpenoids that act as natural reducing and stabilizing agents. UV-Visible spectroscopy confirmed nanoparticle formation with a characteristic absorption peak at 264 nm, while FTIR analysis revealed the involvement of hydroxyl, amine, phenolic, and flavonoid functional groups in nanoparticle stabilization. XRD studies confirmed the crystalline nature of ZnO nanoparticles, and SEM analysis demonstrated predominantly spherical particles with sizes below 30 nm.

The biosynthesized ZnO nanoparticles exhibited significant antibacterial activity against important human pathogens including *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and *Salmonella sp.*, indicating broad-spectrum antimicrobial potential against both Gram-positive and Gram-negative bacteria. Furthermore, the nanoparticles showed promising antioxidant and agricultural properties by enhancing plant growth and reducing environmental stress effects in crop plants. The use of *Cyperus pangorei*, an abundantly available dry grass species with rich bioactive constituents, highlights its biological significance as a sustainable resource for green nanomaterial synthesis. The study demonstrates that *Cyperus pangorei*-mediated ZnO nanoparticles are eco-friendly, cost-effective, and biologically active nanomaterials with potential applications in biomedical, agricultural, and agro-biotechnological fields.

**Keywords:** *Cyperus pangorei*, UV-Vis, ZnO, FTIR, SEM, XRD, *E. coli*.

**Citation:** Rohan Gavankar, Deepa Verma, Ankita S. Tekade, and Virendra Sangode [2026]. Green Synthesis and Characterization of Zinc Oxide Nanoparticles Using *Cyperus pangorei* Grass Extract and its Antibacterial Activity. *Journal of Diversity Studies*.

**DOI:** <https://doi.org/10.51470/JOD.2026.5.1.195>

**Corresponding Author:** Rohan Gavankar, Virendra Sangode

**E-mail Address:** [rohangavankar@vivacollege.org](mailto:rohangavankar@vivacollege.org), [virendrasangode03@gmail.com](mailto:virendrasangode03@gmail.com)

**Article History:** Received 03 February 2026 | Revised 07 March 2026 | Accepted 05 April 2026 | Available Online May 06, 2026

**Copyright:** © 2026 by the author. The license of *Journal of Diversity Studies*. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## Introduction

Green synthesis of nanoparticles using plant extracts has emerged as an ecofriendly, cost-effective, and sustainable approach in nanotechnology. Nanotechnology-based antimicrobial agents have gained considerable attention as promising alternatives to traditional antibiotics. Nanoparticles possess unique antimicrobial mechanisms that enable them to effectively target resistant microorganisms through multiple pathways simultaneously. The utilization of nanoparticles in commercial applications and nanotechnology has seen a sharp rise in attention. Because green synthesis methods employ fewer harmful chemicals, are environmentally friendly, and allow for the one-step synthesis of nanoparticles, their adoption by researchers is quickly increasing.

*Cyperus pangorei* is a perennial sedge belonging to the family Cyperaceae. It is both pantropical & temperature in distribution [5]. Phytochemical investigations of *Cyperus* species have revealed the presence of diverse bioactive secondary metabolites including sesquiterpenoids, coumarins, anthraquinones, flavonoids, terpenoids, cyperones, rotunols,

limonoids, phenolic compounds, and other biologically active constituents [3,6,9]. These phytochemicals possess strong reducing, stabilizing, antioxidant, and antimicrobial properties, making the plant a promising biological resource for green nanoparticle synthesis. Such bioactive metabolites act as natural capping and reducing agents, thereby eliminating the requirement for hazardous chemical reagents commonly used in conventional nanoparticle synthesis methods. Consequently, *Cyperus pangorei* represents an ecofriendly, sustainable, and cost-effective botanical source for the fabrication of biogenic nanoparticles with potential biomedical, antimicrobial, agricultural, and environmental applications. *Cyperus pangorei* plays a crucial role in the bio reduction of metal ions and stabilization of synthesized nanoparticles during the photosynthesis process. Such bioactive metabolites act as natural capping and reducing agents, thereby eliminating the requirement for hazardous chemical reagents commonly used in conventional nanoparticle synthesis methods, metal oxide nanoparticles have been widely employed in medicine can function as catalysts to assist reduce or remove dangerous

substances from the environment, they have applications in the environmental field [4,10]. Metal nanoparticle synthesis has been described using a variety of techniques over the years, including chemical, biological, and physical techniques. There have been reports of the biosynthesis of ZnO NPS in a variety of bacterial and fungal taxa, including *Bacillus subtilis*, *Escherichia coli*, Ureolytic bacteria, and *Lactobacillus plantarum*, as well as in plants like *Aloe vera*, *Sargassum muticum*, *Eichhornia crassipes*, and *Borassus flabellifer* fruit [13]. Numerous researchers have examined the diverse morphologies and noteworthy antibacterial activity of nano-sized ZnO against a broad range of bacterial species [1,2].

The antimicrobial properties of zinc nanoparticles (NPs) indicate their efficacy in combating infections and the essential requirements for using zinc-based nanoparticles as nano-pesticides in farming include both the materials' basic characteristics and the surrounding environment. Comparing nano-pesticides with traditional formulations, the former offers greater stability, superior efficacy, excellent dispersion, and precise release against target species [30].

Every day researchers migrate away from synthetic approaches since green synthesis does not require high pressure, energy, temperature, or harmful substances. The synthesis of nanoparticles through biosynthesis has been offered as an environmentally friendly, low-maintenance, and inexpensive alternative to physical and chemical processes [31]. Sustainable Development Goal (SDG) targets have been attained in part with bimetallic nanoparticles made using green synthesis [22]. Zinc oxide nanoparticles have attracted a lot of attention from researchers nowadays because of their unique properties [21]. These characteristics are potent photosensitivity, increased excitation binding energy, biocompatibility, eco-friendliness, low prices, simplicity of synthesis, thermal conductivity, and resistance to adverse circumstances [17]. Zinc oxide-based nanoproducts can be produced with greater efficiency via the growth of green nanotechnology [18].

## Materials and Methods

### Sample Collection and Preparation of Plant Extract:

Dry Grass samples of *Cyperus pangorei* were collected from vicinity of pond embankments and transferred in a sterile polyethylene bag for further analysis into laboratory. In laboratory the collected plant material thoroughly rinsed in running tap water to remove any physical contaminants and subsequently washed several times with distilled water. 5gm of cleaned air-dried samples was cut into small fragments and thoroughly mixed with 100 ml of distilled water in 250 ml of Erlenmeyer flask. The mixture was maintained at 70°C for 8 min to extract bioactive phytoconstituents responsible for nanoparticle synthesis. The obtained extract was allowed to cool to ambient temperature and filtered through Whatman No. 1 filter paper to remove particulate matter. The clear filtrate was collected and preserved at 4°C for subsequent biosynthesis experiments involving zinc oxide nanoparticles [14,15].

### Biogenic Synthesis of Zinc oxide Nanoparticles :

50 milliliters of distilled water were used to dissolve 1 milligram of zinc acetate, which was then stirred for one hour. Next, 25 mL of plant extract was added to the zinc acetate solution after 20 mL of NaOH solution had been gradually added. After an hour of incubation, the reaction mixture color changed. Three hours were spent with the solution in the stirrer.

The synthesis of ZnO NPs was confirmed by the appearance of yellow color following the incubation period. Centrifugation was used to separate the precipitate from the reaction solution for 15 minutes at 60°C and 8000 rpm. The pellet was then collected. Pellet was kept in airtight vials for future research after being dried for two hours at 80°C in a hot air oven.

### Characterization of Nanoparticles:

#### SEM:

Scanning electron microscope image shows the morphology of the green synthesized ZnO NPS using *Cyperus pangorei*. A Scanning Electron Microscopy (SEM) investigation was performed with a JEOL JSM-6390 model. By depositing a small amount of sample on the copper grid, a thin carbon-coated film was created. Before analysis, the sample film on the SEM grid was dried for 5 minutes under a mercury lamp [26]. The observations revealed that the ZnO NPs were hexagonal in shape and more agglomerated (fig.1).

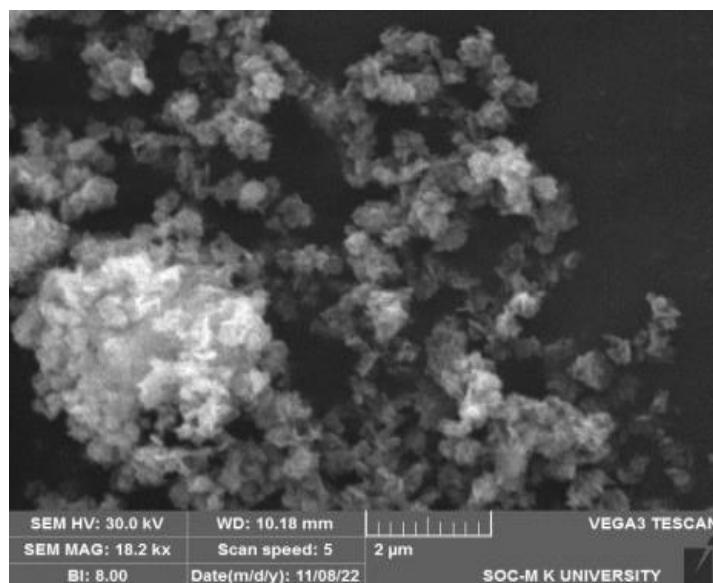


Fig. 1: SEM showing Hexagonal in shape

Utilizing FT-IR spectroscopy, the surface chemistry and involvement of bacterial proteins in the reduction and stability of NPs were investigated. The wavelength spectrum of the cell-free supernatant before and after the addition of metal ion solution ZnONPs was recorded using Perkin Elmer manufacture model spectrum 22575927 RX1 (wavelength range of 4000 cm<sup>-1</sup> to 400 cm<sup>-1</sup>). Blotting paper was used to get rid of any extra water. A small amount of the emulsified sample in the amount of 0.2 was diluted with 10 ml of distilled water for the UV-visible spectrophotometry assessment. This diluted sample was then placed in 1 ml into a 2 ml quartz cuvette. A horizontal slit that was 2 mm broad was placed at the front of the chamber to measure absorption. The UV absorption was further investigated over a spectrum of wavelengths from 100 to 1000 nm. The sample's maximum absorption wavelength was determined to be the wavelength at which the emulsion sample presented its highest level of absorption. The phase purity and particle size of ZnO NPs were determined using the X-ray diffractometer, Bruker AXS (Germany) D8 advanced diffractometer unit. Scherrer's equation (Equation (1)) was used to determine the crystalline size of the synthesized ZnO NPs (Holzwarth et al., 2011).

$$D = k\lambda / \beta \cos \theta$$

$\lambda$ –Is the X-ray wavelength

$\beta$ –Is the Full Width at Half Maximum (FWHM) and  $\theta$ –Is the diffraction angle

$$D = 0.9\lambda / \beta \cos \theta$$

where D is the crystallite size,  $\lambda$  is the wavelength of the x-ray used (1.5406 Å),  $\beta$  is the full width at half maximum (FWHM), and  $\theta$  is the Bragg's angle.

### Characterization

UV spectroscopy and FTIR spectroscopy were applied to zinc oxide nanoparticles. The range of 4000-400cm<sup>-1</sup> was used to record the infrared spectrum. The absorption peak for UV spectroscopic examination was found in each spectrum in the 200–400 nm region, which is a typical band for pure ZnO.

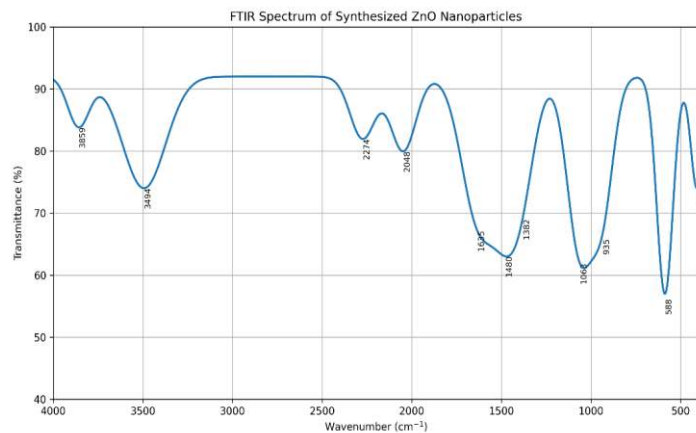


Fig.2 FTIR spectrum of Synthesized ZnO Nanoparticle

No.	Peak	Intensity	Corr. Inte	Base (H)	Base (L)	Area	Corr. Are
1	408.93	1.0055	0.0005	410.86	400.25	21.1888	0.0009
2	588.31	0.5262	0.523	856.43	411.82	943.3609	65.2121
3	935.52	0.9822	0.0567	970.24	857.4	224.0261	1.2172
4	1063.79	0.8938	0.3771	1259.57	971.2	560.614	22.2074
5	1382.06	1.3524	0.1901	1432.21	1260.54	311.9491	4.4975
6	1480.43	1.369	0.1274	1570.12	1433.17	251.6354	2.8475
7	1635.71	1.3569	0.4098	1852.71	1571.09	490.0962	7.869
8	2048.49	2.1284	0.0563	2221.13	1853.67	612.1039	1.9818
9	2274.17	2.1533	0.0192	2337.83	2222.09	192.6494	0.1892
10	2349.4	2.1775	0	2354.22	2338.79	25.6472	0.0001
11	3494.2	0.9387	0.7521	3794.14	2355.19	2629.891	134.7148
12	3859.73	1.5365	0.011	3890.59	3795.11	172.9139	0.1678
13	3973.53	1.4926	0.0156	4000.54	3891.55	198.4032	0.2447

### X-ray Powder Diffraction Analysis

The significant diffraction peaks observed in (010), (002), (011), (012), (110), (013), (020), (112), (021), (004), and (022) planes, and XRD- values represent the average size of 29.34 nm (JCPDS 98-002-7781).

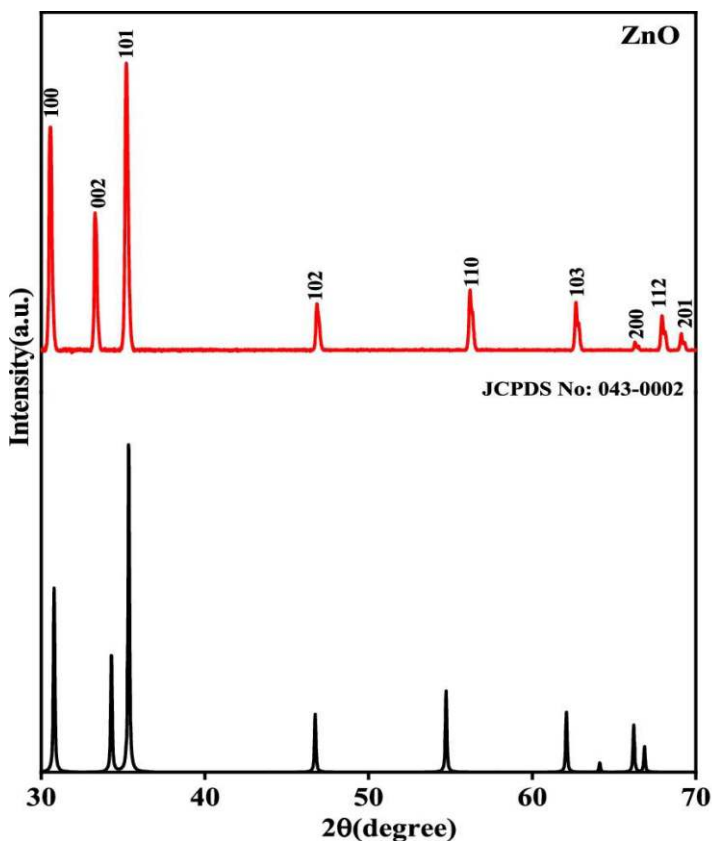


Fig.3: XRD analysis examines the crystalline Phases of green synthesized ZnO Nps.

### Antibacterial Assay of ZnO Nanoparticle:

To evaluate antibacterial potential of biogenic synthesized bacterial strains were used namely *Salmonella sp.*, *Pseudomonas aeruginosa*, *Escherichia coli*, and *Staphylococcus aureus*. The bacterial cultures used in the present investigation were obtained from maintained stock cultures and subculture prior to experimentation to ensure active growth. The antibacterial assay was carried out using the standard disc diffusion technique. Fresh overnight cultures of each bacterial strain were uniformly spread onto sterile nutrient agar plates using sterile cotton swabs under aseptic conditions. Sterile discs impregnated with different concentrations (10 µL and 20 µL) of ZnO nanoparticle suspension were carefully placed on the inoculated agar surface. Commercially available antibiotic discs were used as positive controls for comparison of antibacterial efficacy. The inoculated plates were subsequently incubated at 37°C for 24 h. Following incubation, the antibacterial activity of the synthesized nanoparticles was determined by measuring the diameter of the clear inhibition zones formed around each disc. As a control, commercial antibiotic discs were used. Different degrees of zonation that developed around the disc once the incubation period was over were measured.

### Result and Discussion

The produced zinc oxide nanoparticles; UV-Vis spectra revealed an absorption peak at about 264 nm. The beneficial applications of green synthesized zinc oxide nanoparticles in both agriculture and nanobiotechnology. Earlier reports indicated that ZnO nanoparticles can enhance plant growth and productivity, supporting the plant growth-promoting effects observed in the present study. Similarly, successful biosynthesis of ZnO nanoparticles using plant extracts has been widely reported, where phytochemicals present in botanical materials act as natural reducing and stabilizing agents.

The UV-visible absorption spectra of the biosynthesized ZnO nanoparticles using the leaf extract of *Vernonia cinerea* reveal an absorption peak at 360 nm [32]. functional groups can be identified by analyzing its FT-IR spectra. Higher extract volumes raise the absorbance peak strength, which in turn increases the yield of nanoparticles because the extract contains more phytochemicals. [27]. The synthesis of zinc nitrate to produce zinc oxide nanoparticles is aided by an abundance of hydroxyl (OH) groups present in flavonoids and phenol [29]. Biosynthesized CuO nanoparticles using *Rumex nepalensis* showed a crystalline monoclinic structure with particle sizes ranging from 21–97 nm. The synthesized CuO NPs exhibited significant antimicrobial and antioxidant activities, including effective DPPH free radical scavenging, highlighting their potential biomedical applications and the eco-friendly nature of the green synthesis approach [33]. Green synthesized ZnO nanoparticles using *Nicotiana plumbaginifolia* extract exhibited a hexagonal wurtzite crystalline structure with spherical particles ranging from 16–24 nm. The nanoparticles showed significant antibacterial activity against pathogenic bacteria and strong antioxidant potential with 75.59% DPPH radical scavenging activity, indicating promising biomedical applications [34]. Similar studies were reported in Sustainably synthesized cobalt oxide nanoparticles (CoO NPs) using *Uraria picta* extract demonstrated effective green synthesis through natural reducing and stabilizing phytochemicals. Characterization by UV-Vis, FTIR, XRD, EDS, SEM, and TEM analyses confirmed the structural and morphological properties of the nanoparticles. The biosynthesized CoO NPs exhibited significant antioxidant and antibacterial activities against selected bacterial strains, highlighting their potential biomedical and environmental applications through an eco-friendly and cost-effective synthesis approach [35,37].

The presence of phenolic and flavonoid group molecules plays a role in the inhibition process, stability, and capping agents, while the peak in the 400–600  $\text{cm}^{-1}$  range is designated as Zn–O, suggesting the coordination of produced ZnO nanoparticles [28]. As shown in the SEM image, the synthesized ZnO nanoparticles, with an average particle size of 26 nm estimated by using Debye-Scherrer's formula, revealed efficiency in decreasing the effects of salt stress within *Sorghum bicolor*. The XRD tests examined the particles' hemispherical morphology and sizes below 30 nm. Most of the particles' shapes are spherical and aggregate into larger particles with uncertain shape [24]. *Azadirachta indica* nanoparticles have gained attention in nanotechnology due to their antioxidant and protective properties against reproductive toxicity. Recent studies suggest that neem-mediated nanoparticles may help reduce lead acetate-induced damage in male and female reproductive systems. This review highlights the therapeutic potential of neem-based nanomaterials as eco-friendly alternatives for reproductive health protection [36].

The production of nanoparticles (NPs) from plant extracts has grown into a popular field for research in the past few years. This pattern shows the increased awareness about the potential advantages of green-synthesized metal nanoparticles over the traditional alternatives used in the agricultural sector [20]. Silver nanoparticles are less harmed by zinc nanoparticles (Zn NPs), which have been indicated to be beneficial as a nano fertilizer and an anti-fungal properties agent. *Penicillium expansum*, *Botrytis cinerea*, and *Aspergillus flavus* are just a few of the fungi that zinc nanoparticles are effective against [19,22].

Plenty of nanomaterials help to reduce crop diseases while used in pest management applications. These materials include nano-containers, nano- encapsulates, nano-cages, and nano-emulsion [16]. Zinc oxide nanoparticles, applied to *Vicia faba* seeds extract had significant impacts on *Meloidogyne incognita*, resulting in mortality rates of 88.2%, 93.4%, and 96.72% after exposure times of 24, 48, and 72 hours, in that combination. were treated with five concentrations of ZnO-NPs (12.5, 25, 50, 100, and 200  $\mu\text{g}/\text{mL}$  [25]. The current findings are consistent with these observations, confirming that *Cyperus pangorei* extract can effectively mediate the formation of biologically active ZnO nanoparticles with promising antibacterial and agricultural applications [7,8] According to research published in 2019 by Gomathi and Sathya, zinc nanoparticles exhibit an absorption peak at 277 nm.

The ZnO nanoparticles FT-IR spectra revealed broad bands that occur at 3253.11  $\text{cm}^{-1}$  and correspond to the N-H stretching frequencies of amines. The N=O and C-C stretch is indicated by the bands at 1564.22  $\text{cm}^{-1}$  and 1405.26  $\text{cm}^{-1}$  (Figure 1). The N-O and C-N are seen in the band at 1343.86  $\text{cm}^{-1}$  and 1094.30  $\text{cm}^{-1}$ . The broad bands at 3377.41  $\text{cm}^{-1}$ , which correspond to the O-H and N-H stretching frequencies of alcohol and amine groups, were reported by Myint Khaing et al., in 2018. By evaluating the zone of inhibition, zinc oxide nanoparticles that were manufactured were found to have antibacterial activity against *Salmonella sp.*, *Pseudomonas aeruginosa*, *Escherichia coli*, and *Staphylococcus aureus*. The pathogens were clearly affected by the nanoparticles; anti-bacterial action, as the results plainly revealed. For 10  $\mu\text{l}$  (16mm, 17mm) and 20  $\mu\text{l}$  (17mm, 18mm), the maximum zone of inhibition was shown against *Escherichia coli* and *Pseudomonas aeruginosa*; for 10  $\mu\text{l}$  and 20  $\mu\text{l}$ , the minimum zone of inhibition was seen against *Staphylococcus aureus* (13mm, 13mm) and *Salmonella sp* (15mm, 14mm) (Table 1). The greatest zone of inhibition (32 $\pm$ 0.050) against *Pseudomonas aeruginosa* and the minimum zone of inhibition (25 $\pm$ 0.100) against *Staphylococcus aureus* [11]. The plants branch length grew as the concentration of ZnO nanoparticles rose. The plant growth was higher in the *Cicer arietinum* seeds coated with 0.02g & 0.03g ZnO nanoparticles and the *Vigna radiata* seeds coated with 0.01g ZnO nanoparticles than in the control [12]. Study describes the green synthesis of multiphase iron oxide nanoparticles using *Dolichandrone falcata* leaf extract. The synthesized  $\alpha\text{-Fe}_2\text{O}_3$  nanorods and  $\gamma\text{-Fe}_2\text{O}_3$  spherical nanoparticles were characterized by XRD, FTIR, SEM, and TEM, confirming their crystalline nature with an average size of  $\sim$ 21 nm. The nanoparticles showed significant antibacterial activity, indicating their potential for biomedical and environmental applications, similar study shows green synthesis of nanoparticles using plant extracts is an eco-friendly and cost-effective approach due to the natural reducing and stabilizing agents present in plants. In this study, copper oxide nanoparticles (CuO NPs) were synthesized using *Rivina humilis* L. extract and characterized by UV-Vis, FTIR, XRD, SEM, TEM, and EDS analyses. The biosynthesized CuO NPs exhibited significant antibacterial activity against selected bacterial strains and showed notable antioxidant activity through DPPH free radical scavenging assay. [38,39].

Statistical analysis revealed significant differences ( $p < 0.05$ ) between the positive control and ZnO nanoparticle treatments for most bacterial strains. However, no significant difference was observed between ZnO nanoparticle concentrations of 10  $\mu\text{L}$  and 20  $\mu\text{L}$  in certain bacterial species, indicating comparable antibacterial efficacy at both concentrations.

The enhanced antibacterial activity may be attributed to the small particle size, large surface area, and generation of reactive oxygen species (ROS), which disrupt bacterial cell membranes and interfere with cellular metabolism. These findings demonstrate that green synthesized ZnO nanoparticles using *Cyperus pangorei* possess promising antibacterial potential and could be utilized as eco-friendly antimicrobial agents for biomedical and agricultural applications.

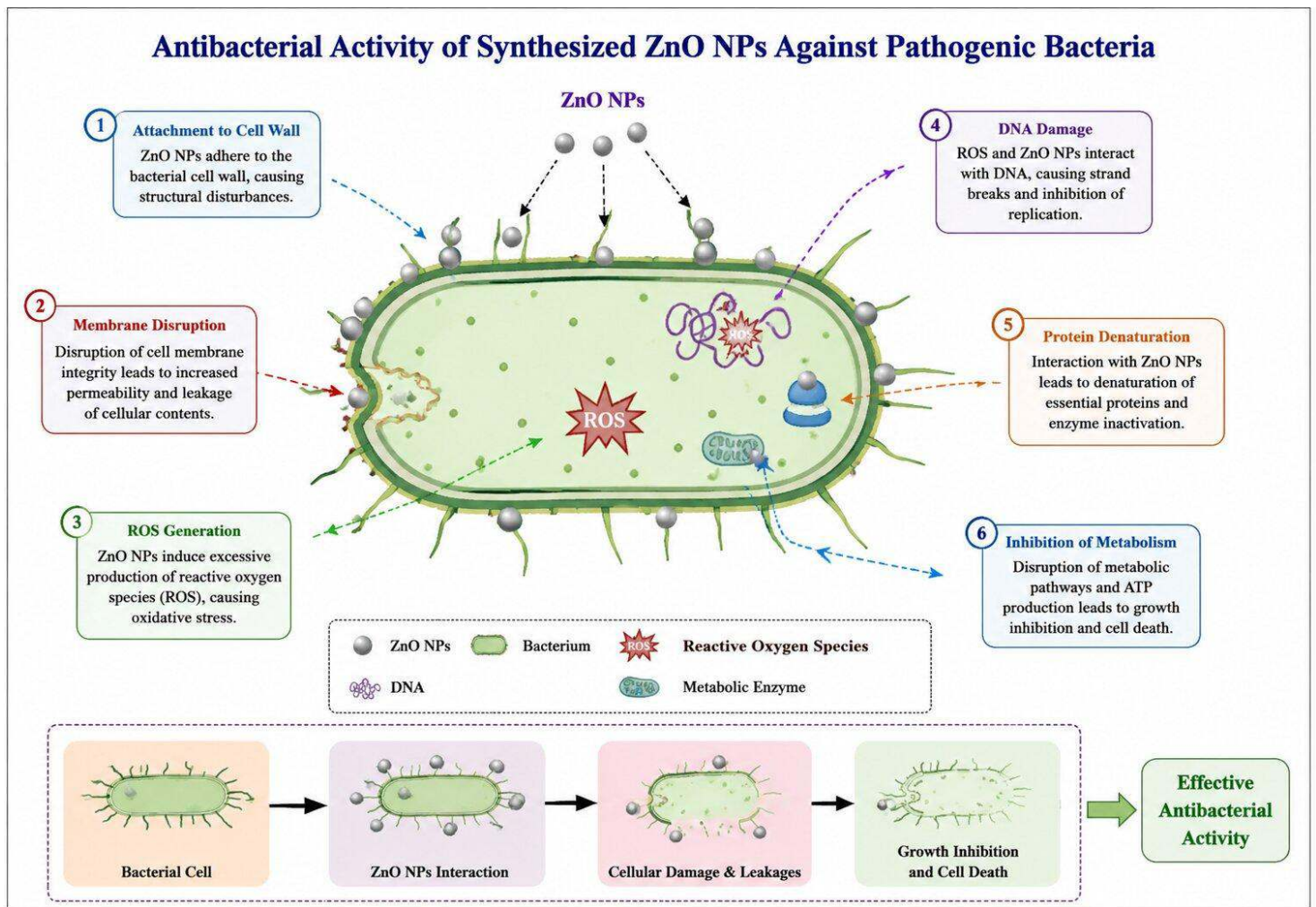


Fig.4: Schematic Representation of Antibacterial activity of Green synthesized ZnO NPS against Pathogenic bacteria

Table 1: Antibacterial Activity of Synthesized ZnO Nps Against Pathogenic Bacteria

S.NO	Microorganism	Zone of Inhibition (mm)		
		Positive Control	ZnO NPs (mm) (10µl)	ZnO NPs (mm) (20µl)
1.	<i>Escherichia coli</i>	25.00 ± 0.58 <sup>a</sup>	16.00 ± 0.33 <sup>b</sup>	17.00 ± 0.58 <sup>b</sup>
2.	<i>Pseudomonas aeruginosa</i>	18.00 ± 0.33 <sup>a</sup>	17.00 ± 0.58 <sup>a</sup>	18.00 ± 0.33 <sup>a</sup>
3.	<i>Salmonella sp</i>	20.00 ± 0.58 <sup>a</sup>	15.00 ± 0.33 <sup>b</sup>	14.00 ± 0.58 <sup>b</sup>
4.	<i>Staphylococcus aureus</i>	22.00 ± 0.33 <sup>a</sup>	13.00 ± 0.58 <sup>b</sup>	13.00 ± 0.33 <sup>b</sup>



Fig. 4: Antibacterial activity of green synthesized ZnO nanoparticles against pathogenic bacteria: (A) *Escherichia coli*, (B) *Pseudomonas aeruginosa*, (C) *Salmonella sp.*, and (D) *Staphylococcus aureus* showing zones of inhibition by agar well diffusion method.

## Conclusion

In the present investigation, an eco-friendly and sustainable approach was employed for the biosynthesis of zinc oxide nanoparticles (ZnO NPs) using dry grass extract of *Cyperus pangorei* as a natural reducing and stabilizing agent. The successful formation of ZnO nanoparticles was confirmed through UV-Visible spectroscopy, which exhibited a characteristic absorption peak, while FTIR analysis revealed the participation of bioactive phytochemicals such as phenolics, flavonoids, hydroxyl, and amine groups in nanoparticle synthesis and stabilization. XRD analysis confirmed the crystalline hexagonal wurtzite structure of ZnO nanoparticles, and SEM micrographs demonstrated predominantly spherical and aggregated nanoparticles with particle sizes below 30 nm. The synthesized ZnO nanoparticles exhibited significant antibacterial activity against both Gram-positive and Gram-negative bacterial pathogens including *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and *Salmonella* sp. The synthesized ZnO nanoparticles exhibited concentration-dependent antibacterial activity against both Gram-positive and Gram-negative bacteria. Statistical analysis revealed significant differences ( $p < 0.05$ ) among treatments, indicating the effectiveness of ZnO nanoparticles against pathogenic bacterial strains. In addition, the nanoparticles showed promising effects on plant growth and stress tolerance, suggesting their potential utility in sustainable agriculture and nano-enabled crop management strategies. The present findings emphasize the biological significance of *Cyperus pangorei* as a rich source of phytochemicals for green nanomaterial synthesis. Compared with conventional chemical methods, the phytochemical synthesis approach offers advantages such as low toxicity, environmental compatibility, cost-effectiveness, and enhanced biological activity, making biosynthesized ZnO nanoparticles promising candidates for biomedical, antimicrobial, and agricultural applications.

## Acknowledgment and Author Contribution:

Plant sample collection and laboratory experiments and discussion write ups were done by Dr. Rohit Gavankar, Dr. Deepa Verma and sample characterization were done by Dr Virendra K Sangode and Dr. Ankita Tekade. Authors wish to Thank Anacon Laboratory Pvt ltd for Characterization of nanomaterial and providing essential lab facilities. The findings of this work demonstrate that *Cyperus pangorei* can serve as a valuable biological source for the production of functional zinc oxide nanoparticles with potential applications in antimicrobial therapy, sustainable agriculture, and other nanobiotechnological fields. The study also contributes to the advancement of green nanotechnology by promoting safer and more ecofriendly nanoparticle fabrication methods.

## References

1. Brayner, R., Ferrari-Iliou, R., Brivois, N., Djediat, S., Benedetti, M. F., & Fiévet, F. (2006). Toxicological impact studies based on *Escherichia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium. *Nano Letters*, 6(4), 866–870. <https://doi.org/10.1021/nl052326h>
2. Buzzea, C., Pacheco, I. I., & Robbie, K. (2007). Nanomaterials and nanoparticles: Sources and toxicity. *Biointerphases*, 2(4), MR17–MR71. <https://doi.org/10.1116/1.2815690>
3. Dini, I., Ramundo, E., Saturnino, P., Scimone, M., & Alcontres, I. S. (1993). Phytochemical investigations on *Cyperus* species. *Biochemical Systematics and Ecology*, 21(2–3), 305–307. [https://doi.org/10.1016/0305-1978\(93\)90079-X](https://doi.org/10.1016/0305-1978(93)90079-X)

4. Gunalan, S., Sivaraj, R., & Rajendran, V. (2012). Green synthesized ZnO nanoparticles against bacterial and fungal pathogens. *Progress in Natural Science: Materials International*, 22(6), 693–700. <https://doi.org/10.1016/j.pnsc.2012.11.015>
5. Haines, R. W., & Lye, K. A. (1993). *The sedges and rushes of East Africa*. Nairobi: East African Natural History Society.
6. Khare, M. D., & Keady, P. (2004). Antimicrobial resistance mechanisms and bacterial adaptation. *Journal of Antimicrobial Chemotherapy*, 53(3), 435–440. <https://doi.org/10.1093/jac/dkh080>
7. Laware, S. L., & Raskar, S. (2014). Influence of zinc oxide nanoparticles on growth, flowering and seed productivity in onion. *International Journal of Current Microbiology and Applied Sciences*, 3(7), 874–881.
8. Khaing, M. M., Thu, M. K., Kyaw, T., Tin, T., & Lwin, T. (2018). Green synthesis of zinc oxide nanoparticles using tropical plants and their characterizations. *International Journal of Scientific & Engineering Research*, 9(8), 551–557.
9. Nyasse, B., Ghogomu, R. T., Sondengam, B. L., Martin, M. T., & Bodo, B. (1988). Chemical constituents from *Cyperus* species. *Phytochemistry*, 27(10), 3319–3321. [https://doi.org/10.1016/0031-9422\(88\)80089-5](https://doi.org/10.1016/0031-9422(88)80089-5)
10. Okitsu, K., Mizukoshi, Y., Yamamoto, T., Maeda, Y., Nagata, Y., & Sonochemical synthesis group. (2007). Sonochemical synthesis of gold nanoparticles on chitosan. *Materials Letters*, 61(17), 3429–3431. <https://doi.org/10.1016/j.matlet.2006.10.039>
11. Kumar Shah, R., Boruah, F., & Parween, N. (2015). Synthesis and characterization of ZnO nanoparticles using leaf extract of *Camellia sinensis* and evaluation of their antimicrobial efficacy. *International Journal of Current Microbiology and Applied Sciences*, 4(8), 444–450.
12. Sundrarajan, M., Ambika, S., & Bharathi, K. (2015). Plant-extract mediated synthesis of ZnO nanoparticles using *Pongamia pinnata* and their activity against pathogenic bacteria. *Advanced Powder Technology*, 26(5), 1294–1299. <https://doi.org/10.1016/j.apt.2015.07.001>
13. Vijayakumar, S., Vinoj, G., Malaikozhundan, B., Shanthi, S., & Vaseeharan, B. (2015). *Plectranthus amboinicus* leaf extract mediated synthesis of zinc oxide nanoparticles and its control of methicillin-resistant *Staphylococcus aureus* biofilm and blood sucking mosquito larvae. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 137, 886–891. <https://doi.org/10.1016/j.saa.2014.08.064>
14. Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2016). A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: A green expertise. *Journal of Advanced Research*, 7(1), 17–28. <https://doi.org/10.1016/j.jare.2015.02.007>
15. Iravani, S. (2011). Green synthesis of metal nanoparticles using plants. *Green Chemistry*, 13(10), 2638–2650. <https://doi.org/10.1039/C1GC15386B>.
16. Abdullah M. Abdo, Amr Fouda, Ahmed M. Eid, Nayer M. Fahmy, Ahmed M. Elsayed, Ahmed Mohamed Aly Khalil, Othman M. Alzahrani, Atef F. Ahmed, and Amal (2022) Eco-Friendly Synthesis and Comparative In Vitro Biological Evaluation of Silver Nanoparticles Using Tagetes erecta Flower Extracts, *Applied Sciences*, 12(2), 887; <https://doi.org/10.3390/app12020887>.
17. Diallo, B.D. Ngom, E.Park, and M.Maaza (2015) Green synthesis of ZnO nanoparticles by *Aspalathus linearis*: Structural & optical properties, *J.of alloys and compounds*, 2015; 646, 425-430, <https://doi.org/10.1016/j.jallcom..05.242>.
18. Gulzar Ahmad Rather, Saima Hamid, Muzafar Riyaz, Musheerul Hassan, Mohammad Ashaq Sofi, Ifrah Manzoor, and Anima Nanda (2022) The Role of Green Synthesised Zinc Oxide Nanoparticles in Agriculture, sustainable agriculture, 11, 119-142.

19. He, L., Liu, Y., Mustapha, A., & Lin, M (2011) Antifungal activity of zinc oxide nanoparticles against *Botrytis cinerea* and *Penicillium expansum*. *Microbiological research*, 166(3): 207-215
20. Jayaseelan, C., Rahuman, A. A., Kirthi, A. V., Marimuthu, S., Santhoshkumar, T., Bagavan, A., & Rao, K. B. (2012) Novel microbial route to synthesize ZnO nanoparticles using *Aeromonas hydrophila* and their activity against pathogenic bacteria and fungi. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 90: 78-84.
21. Khadeeja Parveen, Viktoria Banse, and Lalita Ledwani (2016) Green Synthesis of Nanoparticles: Their Advantages and Disadvantages, *AIP Conference Proceedings*, [1724\(1\) doi:10.1063/1.4945168](https://doi.org/10.1063/1.4945168).
22. Mariana Larranaga Tapia, Benjamin Betancourt -Tovar, Marcelo Videira, Marilena Antunes, and Jorge L. Cholula Diaz (2024) Green synthesis trends and potential applications of bimetallic nanoparticles towards the sustainable development goals 2030, *Royal society of chemistry, nanoscale adv*, 6, 51-71. DOI: [10.1039/D3NA00761H](https://doi.org/10.1039/D3NA00761H)
23. Oluwatoyin Adenike Fabiyi, Ridwan Olamilekan Alabi and Riwan Ali Ansari (2020) Nanoparticles synthesis and their application in the management of Phytonematodes: An overview, *Management of Phytonematodes: Recent Advances and Future Challenges*, Pp 125-140.
24. Padalia H, Moteriya P, Chanda S (2015) Green synthesis of silver nanoparticles from marigold flower and its synergistic antimicrobial potential. *Arab J Chem*, 8(5):732-741.
25. PanelYaqi Jiang, Pingfan Zhou, Peng Zhang, Muhammad Adeel, Noman Shakoor, Yuanbo Li, Mingshu Li, Manlin Guo, Weichen Zhao, Benzhen Lou, Lingqing Wang, Iseult Lyn (2021) Green Synthesis of Zinc Oxide Nanoparticles (ZnO-NPs) by *Pseudomonas aeruginosa* and Their Activity against Pathogenic Microbes and Common House Mosquito, *Culex pipiens*, *MDPI*, 14(22): 6983. doi: [10.3390/ma14226983](https://doi.org/10.3390/ma14226983)
26. Pankaj gupta, and Neeru Vasudeva (2010) *In vitro* antiplasmodial and antimicrobial potential of *Tagetes erecta* roots, *Pharmaceutical Biology*, 28, Pages 1218-1223, <https://doi.org/10.3109/13880201003695142>.
27. Rastogi, J, and Arunachalam (2011) Sunlight based irradiation strategy for rapid green synthesis of highly stable silver nanoparticles using aqueous garlic (*Allium sativum*) extract and their antibacterial potential, *Materials chemistry and physics*, 129, Issue 1-2, 558-563. <https://doi.org/10.1016/j.matchemphys.2011.04.068>.
28. Seifipour, R.; Nozari, M.; Pishkar, L (2021) Preparation of ZnO Nanoparticles Using *Tragopogon Collinus* Leaf Extract and Study of Its Antibacterial Effects for Therapeutic Applications. *J. Plant Biochem. Biotechnol.* 30, 586–595. [Google Scholar] [[CrossRef](#)].
29. Waseem Ahmad, Divya Kalra (2010) Green synthesis, characterization and anti microbial activities of ZnO nanoparticles using *Euphorbia hirta* leaf extract, *Journal of King Saud University-Science*, 32, (4), 2358-2364, <https://doi.org/10.1016/j.jksus.2020.03.014>.
30. Weichen Zhao, Yanwanjing Liu, Peng Zhang, Pingfan Zhou, Zhangguo Wu, Benzhen Lo u, Yaqi Jiang, Noman Shakoor, Mingshu Li, Yuanbo Li, Iseult Lynch, Yukui Rui, Zhiqian g Tan (2022) Engineered Zn-based nano-pesticides as an opportunity for treatment of phytopathogens in agriculture, *Nanoimpact*, Volume 28, <https://doi.org/10.1016/j.impact.2022.100420>
31. Xu, P., He, H., Gao, Q., Zhou, Y., Wu, Z., Zhang, X., Sun, L., Hu, G., Guan, Q., You, Z,X (2021) Human midbrain dopaminergic neuronal differentiation markers predict cell therapy outcome in a Parkinson's disease model. *J. Clin. Invest.* <https://doi.org/10.1172/JCI156768>
32. Zeba Azim, N.B. Singh, Shubhra Khare, Ajey Singh, Nimisha Amidst, Niharika, and Ravi Kumar Yadav (2022) Green synthesis of zinc oxide nanoparticles using *Vernonia cinerea* leaf extract and evaluation as nano-nutrient on the growth and development of tomato seedling, *J.of Plant Nanobiology*, Volume 2, <https://doi.org/10.1016/j.plana.2022.100011>
33. Pawar Abhimanyu , Mungole Arvind , Naktode Kishor Biosynthesis of CuO Nanoparticles Using Plant Extract as a Precursor: Characterization, Antibacterial, and Antioxidant Activity(2023) *Nano Biomedicine and Engineering* 15(4),369-377, <https://doi.org/10.26599/NBE.2023.9290027>.
34. Pawar, A.P., Naktode, K.S., Mungole, A.J. and Anga, S., 2025. Biogenic ZnO nanoparticles: Structural characterisation and bioactivity evaluation. *Chemistry Journal of Moldova*, 20(1), pp.51–61. <https://doi.org/10.19261/cjm.2025.1308>.
35. Sheikh, S., Mungole, A.J., Bhat, A.R., Pawar, A.P., Pandhurnekar, C.P., Pandhurnekar, H.C., Wahab, R., Kanfode, H.P., Upadhye, V.J., Ahmed, S., Parshuramkar, D.M., Al-Omari, M. and Elumalai, P., 2025. *Sustainable plant mediated synthesis of cobalt oxide nanoparticles using Uraria picta extract with enhanced biological activity*. *Scientific Reports*, 15, 44017. <https://doi.org/10.1038/s41598-025-28596-0>
36. Kamal Vatika, Lisha, Shamal Kumbhar, Mehtab Yasmeen, Sunil Akare and Virendra K. Sangode (2024) Nanoparticle of *Azadirachta indica* (Neem) mitigating lead acetate induced reproductive toxicity. *Biochem. Cell. Arch.* 24, 1101-1109. DOI: <https://doi.org/10.51470/bca.2024.24.1.1101>
37. Alok Kumar Srivastava, Virendra Kishor Sangode, K. V. Shalini, and Sangeetha Shanmugam (2026). *Modern Nano-Fertilizer Delivery Systems and Nano-Biofertilizers: Advances, Mechanisms, and Sustainable Applications in Climate-Smart Agriculture: A Comprehensive Review* *Journal: Journal of Diversity Studies* DOI: <https://doi.org/10.51470/JOD.2026.5.1.146>
38. B. G. Yadao, C. P. Pandhurnekar, H. C. Pandhurnekar, R. Pagadala, and A. J. Mungole, "Dolichandrone falcata-Mediated Green Synthesis of Mixed-Phase  $\alpha$ -/ $\gamma$ -Iron Oxide Nanoparticles With Structural Characterization and Antibacterial Properties." *ChemistrySelect* 11, no. 3 (2026): e04313. <https://doi.org/10.1002/slct.202504313>
39. S. Sheikh, A. J. Mungole, C. P. Pandhurnekar, H. P. Kanfode, H. C. Pandhurnekar, A. P. Pawar, *ChemistrySelect* 2024, 9, e202401219. <https://doi.org/10.1002/slct.202401219>